TG1050, an HBV-targeted immunotherapeutics, efficiently decreases HBV viremia and antigenemia in a preclinical model; a meta-analysis and the determination of the involvement of CD4 and CD8 T cells.

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INTRODUCTION

Purpose: Current therapies (nucleos(t)ide analogs or peg-IFNa) for chronic hepatitis B virus (HBV) infection rarely achieve virus clearance. Cohort studies have shown the critical role of cellular immune responses to control HBV infection. We developed an HBV-targeted immunotherapeutic called TG1050 and have shown the induction of multispecific T cells in an HBV-persistent mouse model (AAV-HBV) together with antiviral properties (Martin, Gut 2015 Dec;64(12):1961). We report here a meta-analysis of 5 experiments performed with the AAV-HBV model. Furthermore, we have started to dissect in this HBV model TG1050associated mechanism of action in particular the role of CD4 and CD8 T-cells.

TG1050 : OBJECTIVE AND DESCRIPTION

OBJECTIV

HBV-specific vector-based erapeutic inducing potent. multispecific. cross reactive T cell responses sustained and displaying properties of immune responses detected in HBV resolving patients

TG105 Based on non-replicative E1 and E3 deleted human adenovirus encoding a fusion protei Core fused to a deleted and mutated selected HBsAg domains (genotype D sequence

C	ore t	Pol1	Env 1	Pol2	En
1	148		(37 aa)		(29 :

HBV PERSISTENT MODEL AAV-HBV² (i.v.) <u>Blood</u> AAV-HBV C57BL/6 mice: AAV2: ITR <u>Liver</u> Absence of AAV8: Capsid HBV: 1.2x full length detected **HBV-specific** HBsAg immune responses BeAg C57BL/6

MATERIAL & METHODS

TG1050 treatment in AAV-HBV mice : C57BL/6 mice were injected intravenously with recombinant AAV-HBV, encoding 1.2 copies of the HBV genome. In this model HBV antigens and DNA replication intermediates (HBV mRNA incl. pgRNA) are detected within the liver as well as HBsAg, HBeAg and HBV infectious particles in the blood of injected mice, with a persistence of around 1 year² In parallel no HBV-specific T cells or antibodies were detected in AAV-HBV injected mice whereas an increased number of regulatory T cells and IL-10 producing T cells are present in the liver. This model mimics to some extent the HBV chronic infection. Four to 5 weeks after AAV-HBV injection, TG1050 was injected multiple times by sub-cutaneous route at a dose of 2x10⁹ vp/injection/mice and viral parameters were monitored at various time points. HBsAg in mouse sera was quantified using a BioRad kit (Monolisa HBsAg plus using a recombinant HBsAg protein for quantification). HBV DNA in mouse sera was quantified using a qPCR assay (limit of quantification : 100 copies/reaction). Anti-HBsAg/HBcAg antibodies were detected by an « in house » ELISA assay.

Meta-analysis: For the meta-analysis 5 preclinical experiments, lasting in total between 11 weeks to 20 weeks, were considered for statistical analyses: Female mice, C57BL/6 strain, AAV only or AAV+TG1050 treatment (at 2x10⁹ vp/injection/mice), at least 3 time points after TG1050 administration.

Global Mixed model: A global mixed model was done with all experiments considering the following covariates: Time, Treatment, the interaction between Time and Treatment and the HBsAg value at baseline as fixed effects and the Experiment as random effect.

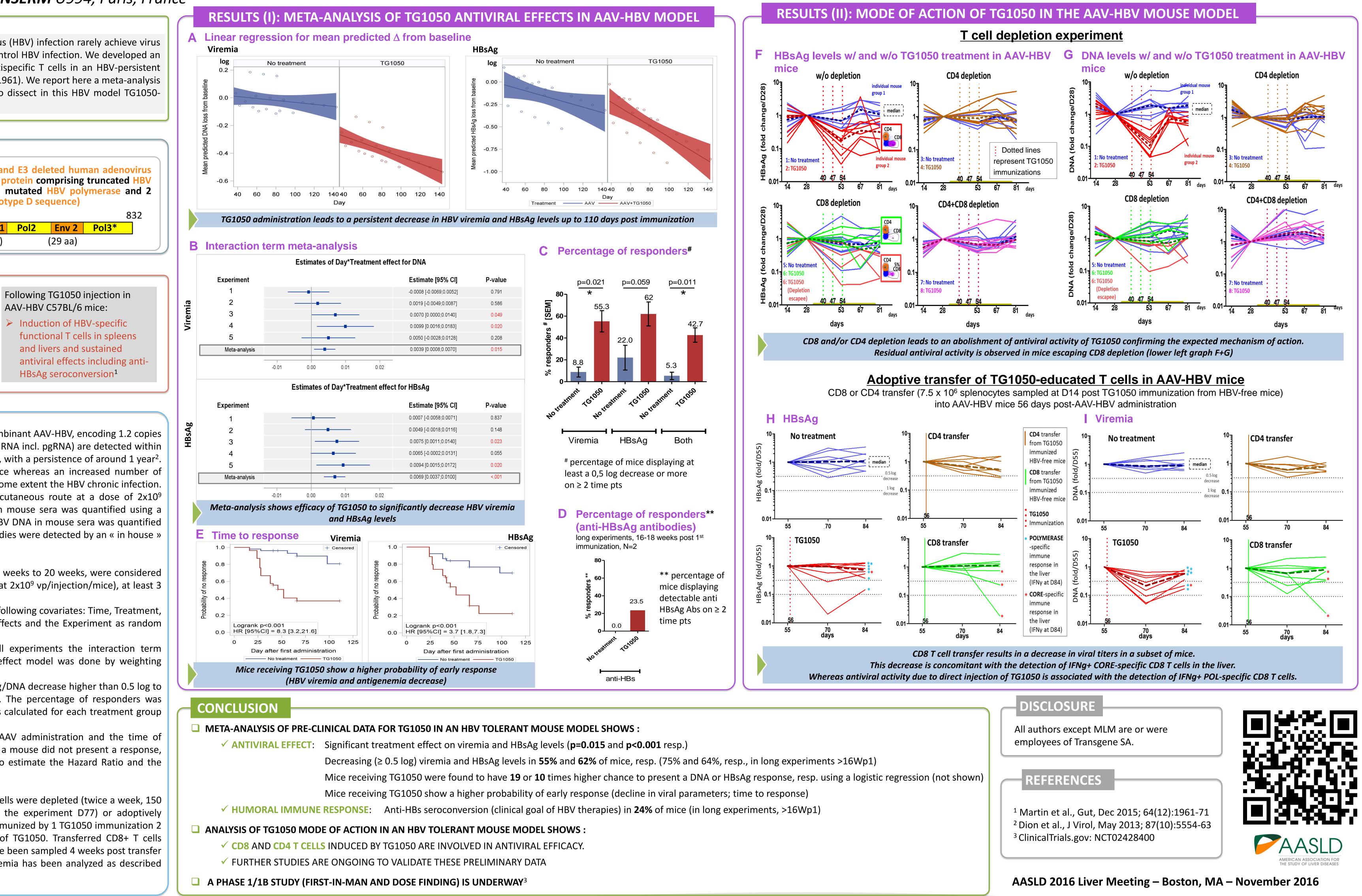
Meta-Analysis for interaction term: The meta-analysis was done by estimating for all experiments the interaction term Day*Treatment and the associated standard error. Then a meta-analysis using a fixed effect model was done by weighting estimation with the inverse-variance.

Percentage of responder: A mouse was considered as "Responder" if it presented an HBsAg/DNA decrease higher than 0.5 log to the baseline value for two or more time points during the study (consecutive or not). The percentage of responders was calculated for each experiment and then the mean average percentage of responders was calculated for each treatment group and compared with a non-parametric Wilcoxon-Mann-Whitney test.

TTR: The Time-To-Response (TTR) was defined as the time between the first TG1050/AAV administration and the time of response (defined as the second time point presenting a decreased more than 0.5 log). If a mouse did not present a response, the TTR was censored at the last blood sample measurement. A Cox model was done to estimate the Hazard Ratio and the estimated confidence interval.

TG1050 mode of action experiments : In similar experimental settings, CD4 and/or CD8 T cells were depleted (twice a week, 150 μg, starting at D34 before 3 TG1050 immunizations (D40, D47, D54) until the end of the experiment D77) or adoptively transferred (7.5x10⁶ CD8+ or CD4+ cells purified by magnetic beads from HBV-free mice immunized by 1 TG1050 immunization 2 weeks before into AAV-HBV mice) to determine their involvement in antiviral efficacy of TG1050. Transferred CD8+ T cells contained ~1-2x10⁵ HBV-specific (polymerase and core) multimer positive T cells. Livers have been sampled 4 weeks post transfer to analyze intracellular IFNg secretion by intrahepatic CD8 T cells. (Serum HBsAg and viremia has been analyzed as described before.)





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